(FILE 'HOME' ENTERED AT 11:53:59 ON 12 JUN 2002)

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FILE 'MEDLINE, AGRICOLA, CANCERLIT, SCISEARCH, CAPLUS, BIOSIS, MEDICONF'
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        1659511 S FIBROBLAST? OR EPIDER? OR SKIN OR DERMIS OR DERMAL
L1
          85661 S L1 AND (GRAFT OR TRANSPLANT?)
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           3666 S L2 AND ((EXTRACELULAR (S) MATRIX) OR DECORIN OR COLLAGEN OR
L3
           3666 FOCUS L3 1-
L4
                 E MURPHY MICHAEL?/AU
                 E MURPHY MIC?/AU
             126 S E30
L5
               8 S L1 AND L5
L6
               5 DUP REM L6 (3 DUPLICATES REMOVED)
1.7
               5 SORT L7 PY
L8
L9
              0 S L3 AND L8
              0 S L2 AND L8
L10
              14 S L4 AND BIOENGINEE?
L11
              9 DUP REM L11 (5 DUPLICATES REMOVED)
L12
               9 SORT L12 PY
L13
             165 S L2 AND (TISSUE (S) CONSTRUCT)
             100 DUP REM L14 (65 DUPLICATES REMOVED)
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             100 FOCUS L15 1-
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     ANSWER 4 OF 5 CAPLUS COPYRIGHT 2002 ACS
L8
     2000:351643 CAPLUS
AN
DN
     132:331698
     Bioengineered tissue constructs and methods for producing and using them
TT
     PCT Int. Appl., 68 pp.
     CODEN: PIXXD2
     Murphy, Michael P.; Ronfard, Vincent
IN
     Cultured tissue constructs comprising cultured cells and endogenously
AB
     produced extracellular matrix components without the requirement of
     exogenous matrix components or network support or scaffold members. Some
     tissue constructs of the invention are comprised of multiple cell layers
     or more than one cell type. The tissue constructs of the invention have
     morphol. features and functions similar to tissues and their strength
     makes them easily handleable. Preferred cultured tissue constructs of the
     invention are prepd. in defined media, i.e., without the addn. of chem.
     undefined components.
                                              APPLICATION NO. DATE
     PATENT NO.
                       KIND DATE
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                       A1 20000525
                                             WO 1999-US27505 19991119
     WO 2000029553
PΙ
          W: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE,
              DK, EE, ES, FI, GB, GE, GH, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM
          RW: GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW, AT, BE, CH, CY, DE,
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              CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG
     EP 1131410
                        A1 20010912
                                             EP 1999-962807 19991119
             AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT,
              IE, FI
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Basement membrane Culture media Extracellular matrix Fibril Fibroblast Intestine Lung Skin Tendon Umbilical cord Urethra (bioengineered tissue constructs and methods for producing and using them) Collagens, biological studies IT Decorins Glycosaminoglycans, biological studies Tenascins RL: BPN (Biosynthetic preparation); BSU (Biological study, unclassified); BIOL (Biological study); PREP (Preparation) (bioengineered tissue constructs and methods for producing and using them) ΙT Growth factors, animal RL: BPR (Biological process); BSU (Biological study, unclassified); BIOL (Biological study); PROC (Process) (bioengineered tissue constructs and methods for producing and using Hormones, animal, biological studies ΙT RL: BPR (Biological process); BSU (Biological study, unclassified); BIOL (Biological study); PROC (Process) (bioengineered tissue constructs and methods for producing and using them) Peptides, biological studies IT RL: BPR (Biological process); BSU (Biological study, unclassified); BIOL (Biological study); PROC (Process) (bioengineered tissue constructs and methods for producing and using them) Proteins, general, biological studies IT RL: BPR (Biological process); BSU (Biological study, unclassified); BIOL (Biological study); PROC (Process) (bioengineered tissue constructs and methods for producing and using them) IT RL: BSU (Biological study, unclassified); BIOL (Biological study) (bioengineered tissue constructs and methods for producing and using them) IT Eye (cornea, epithelium; bioengineered tissue constructs and methods for producing and using them) IT Eye (cornea, stroma; bioengineered tissue constructs and methods for producing and using them) IT Skin (dermis, papilla; bioengineered tissue constructs and methods for producing and using them) TΤ (dermis; bioengineered tissue constructs and methods for producing and using them) IT (epidermis; bioengineered tissue constructs and methods for producing and using them) IT Bladder Esophagus (epithelium; bioengineered tissue constructs and methods for producing and using them) IT Hair (follicles; bioengineered tissue constructs and methods for producing and using them) ΙT (keratinocyte; bioengineered tissue constructs and methods for producing and using them)

Mouth (mucosa, epithelium; bioengineered tissue constructs and methods for producing and using them) IT Mouth (mucosa; bioengineered tissue constructs and methods for producing and using them) IT Penis (prepuce, Neonate male; bioengineered tissue constructs and methods for producing and using them) IT Skin (stratum corneum; bioengineered tissue constructs and methods for producing and using them) Collagens, biological studies IT RL: BPN (Biosynthetic preparation); BSU (Biological study, unclassified); BIOL (Biological study); PREP (Preparation) (type I; bioengineered tissue constructs and methods for producing and using them) Collagens, biological studies IT RL: BPN (Biosynthetic preparation); BSU (Biological study, unclassified); BIOL (Biological study); PREP (Preparation) (type III; bioengineered tissue constructs and methods for producing and using them) THERE ARE 4 CITED REFERENCES AVAILABLE FOR THIS RECORD RE.CNT 4 RE (1) Bell, E; US 5536656 A 1996 (2) Cohen; ANNALS OF BIOMEDICAL ENGINEERING 1991, V19(5), P600

(3) Organogenesis Inc; WO 9531473 A 1995 CAPLUS

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(4) Takeda Chemical Industries; EP 0282746 A 1988 CAPLUS

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(FILE 'HOME' ENTERED AT 11:53:59 ON 12 JUN 2002) FILE 'MEDLINE, AGRICOLA, CANCERLIT, SCISEARCH, CAPLUS, BIOSIS, MEDICONF' ENTERED AT 11:54:07 ON 12 JUN 2002 1659511 S FIBROBLAST? OR EPIDER? OR SKIN OR DERMIS OR DERMAL L185661 S L1 AND (GRAFT OR TRANSPLANT?) L23666 S L2 AND ((EXTRACELULAR (S) MATRIX) OR DECORIN OR COLLAGEN OR L3 3666 FOCUS L3 1-L4=> d an ti so au ab pi 14 4 9 10 ANSWER 4 OF 3666 CAPLUS COPYRIGHT 2002 ACS T.4 1990:104894 CAPLUS AN 112:104894 DNEpidermal graft system containing collagen TI -coated surgical dressing PCT Int. Appl., 45 pp. SO CODEN: PIXXD2 Brysk, Miriam M. IN A skin autograft or allograft composite is prepd. by culturing AB epidermal cells in a medium low in Ca2+ which prevents cell differentiation, layering the cells on a sheet of collagen -coated pliable material such as a synthetic surgical dressing, inverting the sheet on a recipient, and allowing it to remain in position until epidermal cells attach to the recipient and facilitate the formation of a skinlike covering. Thus, 1 of the 2 polyethylene surface layers of a Vigilon polyethylene/hydrocolloid-type surgical dressing was removed and the dressing was coated with a collagen soln. and dried. Trypsin-dissocd. cells from human epidermis sections were cultured and subcultured to .apprx.75% confluence in MCBD-153 medium having a low Ca2+-concn. and seeded onto the collagen-coated surgical dressing; after 4-6 h, the dressing was inverted onto a wound. KIND DATE APPLICATION NO. DATE PATENT NO. -------------\_ \_ \_ \_ -----WO 1988-US3602 19881014 WO 8903228 A1 19890420 PΤ W: AT, AU, BB, BG, BR, CH, DE, DK, FI, GB, HU, JP, KP, KR, LK, LU, MC, MG, MW, NL, NO, RO, SD, SE, SU RW: AT, BE, BJ, CF, CG, CH, CM, DE, FR, GA, GB, IT, LU, ML, MR, NL, SE, SN, TD, TG 19880209 19910514 US 1988-153957 US 5015584 Α 19881014 19890502 AU 1988-26259 AU 8826259 A1 19940802 US 1991-672840 19910321 US 5334527 Α ANSWER 9 OF 3666 CAPLUS COPYRIGHT 2002 ACS L41979:70310 CAPLUS AN 90:70310 DN Mechanism of skin graft adherence: collagen , elastin, and fibrin interactions SO Surg. Forum (1977), 28, 522-4 CODEN: SUFOAX; ISSN: 0071-8041 Tavis, Michael J.; Thornton, James W.; Harney, John H.; Danet, Richard T.; AU Woodroof, Aubrey; Bartlett, Robert H. After 72 h of graft placement, collagen grafts AB demonstrated a mean adherence to exposed deep fascia on rats equiv. to that of skin autografts, and both of these were significantly more adherent than elastin grafts. Transmission electron micrographs of the graft-wound interface indicated a surface interaction between collagen in the graft and fibrin, and fibrin may be the binding protein in graft adherence. Grafts pretreated with heparin prior to placement showed a significant redn. in adherence, and similar results were obtained when fibrinolysin was applied to the surface of intact grafts. Collagen-specific fibrin binding was also demonstrated when collagen disks were exposed to 125I-labeled fibrinogen. The fibrinogen binding kinetics with both collagen and elastin closely correlated with in vivo adherence values. Thus, grafts are apparently bound to wounds during the initial 72 h of placement by

fibrin, and this process may involve surface interactions between fibrin

and collagen rather than elastin.

L4 ANSWER 10 OF 3666 CAPLUS COPYRIGHT 2002 ACS AN 2000:90446 CAPLUS DN 133:86409

TI Methods for the serum-free culture of keratinocytes and transplantation of collagen-GAG-based skin substitutes

Methods in Molecular Medicine (1999), 18(Tissue Engineering Methods and Protocols), 365-389
CODEN: MMMEFN

AU Boyce, Steven T.

AB This chapter describes specific techniques for prepn. and grafting to surgical wounds in athymic mice of cultured **skin** substitutes from **collagen**-glycosaminoglycan substrates populated with normal human keratinocytes, melanocytes, and **fibroblasts** grown in serum-free or low-serum conditions.

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          85661 S L1 AND (GRAFT OR TRANSPLANT?)
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           3666 S L2 AND ((EXTRACELULAR (S) MATRIX) OR DECORIN OR COLLAGEN OR
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           3666 FOCUS L3 1-
                 E MURPHY MICHAEL?/AU
                 E MURPHY MIC?/AU
            126 S E30
L5
              8 S L1 AND L5
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              5 DUP REM L6 (3 DUPLICATES REMOVED)
L7
              5 SORT L7 PY
              0 S L3 AND L8
L9
              0 S L2 AND L8
L10
             14 S L4 AND BIOENGINEE?
L11
              9 DUP REM L11 (5 DUPLICATES REMOVED)
L12
              9 SORT L12 PY
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L13 ANSWER 4 OF 9
                        MEDLINE
                     MEDLINE
     1998184226
     Genetically modified human keratinocytes overexpressing PDGF-A enhance the
     performance of a composite skin graft.
     HUMAN GENE THERAPY, (1998 Mar 1) 9 (4) 529-39.
     Journal code: 9008950. ISSN: 1043-0342.
     Eming S A; Medalie D A; Tompkins R G; Yarmush M L; Morgan J R
     Skin loss due to burns and ulcers is a major medical problem.
AB
     Bioengineered skin substitutes that use cultured
     keratinocytes as an epidermal layer with or without analogues of
     the dermis are one strategy for skin repair. However,
     none can achieve definitive wound closure, function, or cosmesis
     comparable to split-thickness autografts. Moreover, autograft donor sites,
     which require time to heal, may be limited or have attendant problems such as infection or functional/cosmetic deficiencies. To determine if the
     performance of composite {\it skin grafts} of keratinocytes
     on a dermal analogue could be enhanced, human keratinocytes were
     genetically modified to overexpress platelet-derived growth factor A chain
     (PDGF-A). Composite grafts of modified keratinocytes seeded onto
     acellular dermis, prepared from cryopreserved cadaver skin, secreted PDGF-AA protein in vitro [90 ng/graft
     (1.5 \times 1.5 \text{ cm})/24 \text{ hr}]. To test their performance in a wound healing model,
     composite grafts were transplanted to full-thickness
     excisional wounds on the back of athymic mice. PDGF-A grafts
     formed a stratified differentiated epidermis similar to control
     grafts. The acellular dermis was repopulated with host
     fibrovascular cells and by day 7, the PDGF-A grafts had
     significantly more cells in the dermis and increased staining
     for murine collagen types I and IV. At this early time point,
     wound contraction was also significantly inhibited in PDGF-A
     grafts versus control grafts. Thus, PDGF-A
     overexpression improves graft performance during the first
     critical week after transplantation.
L13 ANSWER 5 OF 9 CAPLUS COPYRIGHT 2002 ACS
     1999:405070 CAPLUS
DN
     131:63508
     Chondrocyte-like cells useful for tissue engineering and methods
SO
     PCT Int. Appl., 31 pp.
     CODEN: PIXXD2
     Bhatnagar, Rajendra S.; Nicoll, Steven B.
IN
     Fibroblast cells are treated with a chem. inhibitor of protein
AB
     kinase C such as staurosporine, in conjunction with functionally hypoxic
     micromass culture so as to be induced into chondrogenic differentiation.
     Such fibroblast-derived, chondrocyte-like cells may be seeded
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onto three-dimensional polymer scaffolds for use in the repair of articular cartilage lesions, and thus can obviate the need for invasive

techniques to harvest autologous chondrocytes from a limited supply of existing articular cartilage, or to avoid the need for obtaining allogenetic chondrocytes from non-biocompatible donor tissues.

	PATENT NO.	KIND DATE	APPLICATION NO.	DATE
ΡI	WO 9931221	A1 19990624	WO 1998-US25918	19981207
	W: AL, AM,	AT, AU, AZ, BA, BB	, BG, BR, BY, CA, CH	, CN, CU, CZ, DE,
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	MX, NO,	NZ, PL, PT, RO, RU	, SD, SE, SG, SI, SK	, SL, TJ, TM, TR,
	TT, UA,	UG, US, UZ, VN, YU	, ZW, AM, AZ, BY, KG	, KZ, MD, RU, TJ, TM
			, UG, ZW, AT, BE, CH	
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		GN, GW, ML, MR, NE		
	US 6197586	B1 20010306	US 1998-204824	19981203
	CA 2313808		CA 1998-2313808	19981207
	AU 9916296	A1 19990705	AU 1999-16296	19981207
			EP 1998-960780	
			, GB, GR, IT, LI, LU	
	IE, FI			
		A1 20010628	US 2001-760629	20010116

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migration, a porous construct contg. cells to be transplanted, and (b) delivering the construct into an area of interest in the living subject to form a vascularized three-dimensional tissue. The preferred construct has a dimension in which it is about 50 .mu.mm to about 500 .mu.mm from the outermost surface to the center of the construct. The preferred construct also has an interconnected porous structure having a pore size of from about 10 .mu.mm to no greater than 300 .mu.mm. The cells within the preferred construct are no greater than 250 .mu.mm from an outer surface of the construct.

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PATENT NO.
                 KIND DATE
                                                 APPLICATION NO. DATE
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WO 9952356 A1 19991021
                                              WO 1999-US7816 19990409
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          TR, TT, UA, UG, US, UZ, VN, YU, ZW, AM, AZ, BY, KG, KZ, MD, RU,
          TJ, TM
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                       A1
AU 9935520
                              19991101
                                                 AU 1999-35520
     R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, FI
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JP 2002511284
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                              20020416
                                                 JP 2000-542979
                                                                        19990409
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L16 ANSWER 9 OF 100 MEDLINE

AN 2001676206 MEDLINE

TI Tissue-engineered skin. Current status in wound healing.

SO Am J Clin Dermatol, (2001) 2 (5) 305-13. Ref: 64

Journal code: 100895290. ISSN: 1175-0561.

AU Bello Y M; Falabella A F; Eaglstein W H

Tissue-engineered skin is a significant advance in the field of wound healing and was developed due to limitations associated with the use of autografts. These limitations include the creation of a donor site which is at risk of developing pain, scarring, infection and/or slow healing. A number of products are commercially available and many others are in development. Cultured <code>epidermal</code> autografts can provide permanent coverage of large area from a skin biopsy. However, 3 weeks are needed for graft cultivation. Cultured epidermal allografts are available immediately and no biopsy is necessary. They can be cryopreserved and banked, but are not currently commercially available. A nonliving allogeneic acellular dermal matrix with intact basement membrane complex (Alloderm) is immunologically inert. It prepares the wound bed for grafting allowing improved cultured allograft 'take' and provides an intact basement membrane. A nonliving extracellular matrix of collagen and chondroitin-6-sulfate with silicone backing (Integra) serves to generate neodermis. A collagen and glycosaminoglycan dermal matrix inoculated with autologous fibroblasts and keratinocytes has been investigated but is not commercially available. It requires 3 to 4 weeks for cultivation. Dermagraft consists of living allogeneic dermal fibroblasts grown on degradable scaffold. It has good resistance to tearing.  $\bar{\mathbf{A}}\mathbf{n}$  extracellular matrix generated by allogeneic human dermal fibroblasts (TransCyte) serves as a matrix for neodermis generation. Apligraf is a living allogeneic bilayered construct containing keratinocytes, fibroblasts and bovine type I collagen. It can be used on an outpatient basis and avoids the need for a donor site wound. Another living skin equivalent, composite cultured skin (OrCel), consists of allogeneic fibroblasts and keratinocytes seeded on opposite sides of bilayered matrix of bovine collagen. There are limited clinical data available for this product, but large clinical trials are ongoing. Limited data are also available for 2 types of dressing material derived from pigs: porcine small intestinal submucosa acellular collagen matrix (Oasis) and an acellular xenogeneic collagen matrix (E-Z-Derm). Both products have a long shelf life. Other novel skin substitutes are being

investigated. The potential risks and benefits of using **tissue** -engineered **skin** need to be further evaluated in clinical trials but it is obvious that they offer a new option for the treatment of wounds.